



Bioscience Technology Facility

**Flexible access options, cutting-edge
technology and expert guidance for
unparalleled research impact**



UNIVERSITY
of York



Who we are and what we do



The Bioscience Technology Facility at the University of York provides **flexible access** to **cutting-edge technology** and **expert research** support for both academic and industry partners. Our clients include research groups from within our own university, universities across the world and an array of commercial enterprises. Our unique facility capabilities and special **focus on knowledge transfer** allows us to offer **bespoke research solutions** that drive forward projects efficiently and cost effectively. Whether you're exploring unknown realms of scientific enquiry, innovating novel health technologies, require advanced practitioner training or product development and testing, we can assist you in achieving your objectives and **accelerate transformation**.

Peter O'Toole

Director of the Bioscience Technology Facility
and President of the Royal Microscopical Society
(2023-2026)



Explore our offering

The Technology Facility provides an integrated package of technologies and expertise to drive forward your research and development.

Contents:

4

Our facility

5

Service access options

6

Our laboratory leaders

Laboratories:

7

Imaging and Cytometry

10

Genomics

12

Metabolomics and Proteomics

15

Molecular Interactions

16

Protein Production

18

Data Science

20

Our facility in numbers

Our facility

Flexible access options, cutting-edge technology and expert guidance for unparalleled research impact



**6
labs**



**30
staff**



**Over £8 million
invested since 2018**



**Over 80 expertly maintained
and regularly serviced
high-end instruments**



**Over 8,000
people trained
since inception**



**Significant industry
engagement with leading
manufacturers**



**Network of
collaboration covering
over 20 countries**



**Emerging technology
hub with access to
breakthrough innovations**

'The Technology Facility staff were proactive and flexible. Excellent communication was maintained throughout our diverse collaborations. Each experimental step was discussed and met with high quality advice and enthusiasm. I highly recommend working with the York Technology facility!'

**Dr Heiko Wurdak, University of Leeds and
Prof Hrvoje Miletic, University of Bergen**

Service access options

The Technology Facility is open to both external academic and commercial users, and we have a range of access options to suit every need

Direct usage

Most of our equipment is available for use by individuals who, after training, can access the instrumentation independently, with support from staff when needed. Office facilities are also available.

- **Access** to state-of-the-art equipment – when you need it.



Full service

We can provide a full service, with our experienced staff undertaking your experiments for you.

- **Engage** our expert staff to optimise and execute your experiments.



Consultancy and collaborative research

We can help you develop your research by providing expert consultancy and working collaboratively with you.

- **Collaborate** with us to develop your research.



Training and courses

We offer a range of flexible training opportunities and can host extended visits for your staff, allowing them to work alongside our experts.

- **Learn** with us to develop your team.



Charging

Pay-as-you-go (pay for the hours or runs used) or receive a quote for a set piece of work.

Quality management

We can engage our Quality Management System that can be adapted to suit your requirements. Formal auditing by your internal QA department can be arranged as needed.

Our laboratory leaders



Prof Peter O'Toole
Imaging and
Cytometry

Research focus: Technology and method development

- Over 20 years experience in delivering microscopy and cytometry training courses
- President of the Royal Microscopical Society, executive committee member of CTLS, steering committee member of the European Light Microscopy Initiative
- Consultant for multiple large international companies and national small-medium enterprises



Dr Andrew Leech
Molecular
Interactions

Research focus: Biophysical and spectroscopic techniques

- Over 20 years experience in the characterisation of proteins and other biomolecules and their interactions, using labelled and unlabelled, surface-bound and free solution techniques
- Expert in applying spectrophotometric and interaction technologies to investigate protein-protein interactions, protein-ligand binding, and macromolecular stability



Dr Sally James
Genomics

Research focus: Genomic technologies and sequencing method development

- Over 20 years laboratory experience, and over 10 years in developing genomics methods
- Early adopter of long-read nanopore sequencing technology
- Extensive experience in diverse technologies, including long- and short-read sequencing, gene expression, single-cell, and spatial transcriptomics, applied across a wide range of biological systems



Dr Jared Cartwright
Protein Production

Research focus: Expression, purification and characterisation of recombinant proteins

- Established protein production experience in prokaryotic and eukaryotic systems
- Extensive hands-on experience in secreted protein production and optimising yields across *Pichia pastoris*, mammalian cell and baculovirus-insect cell systems
- Fermentation-scale production capabilities



Dr Tony Larson
Metabolomics
and Proteomics

Research focus: Mass spectrometry technologies

- Extensive experience in securing investment in state-of-the-art mass spectrometry platforms for proteomic, metabolomic and lipidomic workflows
- Over 20 years experience in method development and bespoke software workflows for untargeted small molecule, drug, and lipid analysis from complex datasets



Dr Alastair Droop
Data Science

Research focus: Bioinformatics, computational biology & visualisation

- Over 15 years experience in bioinformatics and computational biology
- Extensive expertise in bespoke software design and implementation across multiple languages (R, Python and Rust)
- Prior small-medium enterprise experience running a bioinformatics consultancy company

Imaging and Cytometry

The Imaging and Cytometry Laboratory is committed to bringing state-of-the-art technology to researchers of all skill levels. We devote significant effort to training and knowledge transfer.

Flow cytometry

We offer single-cell analysis and sorting technologies, supported by expertise covering a wide range of sample types. Our state-of-the-art flow cytometers are housed within temperature-controlled Category 2 rooms seamlessly integrated with our core 'omics labs. These include spectral and imaging sorters.

Advanced light microscopy

Explore a comprehensive range of advanced light microscopy technologies, from standard confocal and multiphoton microscopes, to super-resolution and label-free methods. We have expertise in many biophysical applications (e.g. FCS, FCCS, FRAP, FRET, PALM, STORM, SIM) and sample types (organic and inorganic).

Electron microscopy

We have the latest volume STEM capable of both transmission electron microscope (TEM) and scanning electron microscope (SEM), 3D volume EM, EDAX, CLEM and much more, along with a range of peripheral equipment.

Label-free imaging

We are early adopters of quantitative phase imaging techniques such as ptychography for prolonged time-lapse and screening in multi-well plates, as well as digital holography for rapid 3D super-resolution, label-free imaging. We have both Tomocube and Phasefocus options available.

Slide scanning (automated, 100 slide capacity)

We can perform whole slide digitisation, brightfield and multi-channel fluorescence imaging.

Prof Peter O'Toole



Head of Imaging
and Cytometry

✉ peter.otoole@york.ac.uk

☎ 01904 328722

📍 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD

Spatial-omics

The Imaging and Cytometry, Genomics and Data Science laboratories work together to support a range of spatial analysis platforms as part of full service or collaborative projects, using both 10X Genomics and Nanostring systems.

Peripheral equipment and histology

We provide a full service for most types of specimen.

Analytic software packages

We have a range of offline analysis packages for image analysis and flow data analysis.

Imaging and Cytometry in action

Case study

The user need

- Improved methods to quantify the levels of target membrane proteins in a range of cell types.
- Design of a novel rapid flow cytometric assay.

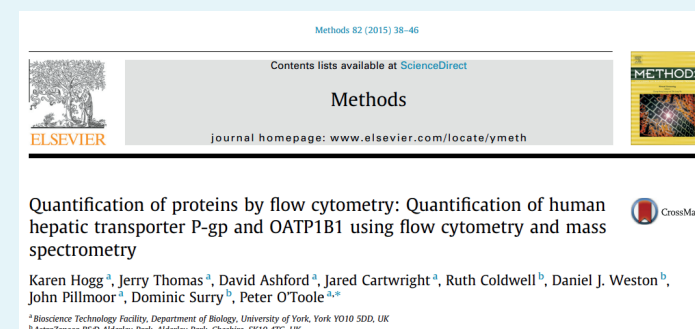
This was a complex longer-term project centred around the Imaging and Cytometry Labs. The workstream utilised our entire breadth of expertise and instrumentation types at York and needed careful integration of resource across the Facility in close collaboration with AstraZeneca's own experts.

First, we needed to establish a range of cell cultures and primary human tissues. Following this, we developed a new antibody design and characterisation process, alongside the production of labelled protein standards together with our Protein Production and Metabolomics and Proteomics laboratories.

The assay we developed was then compared with industry-standard mass spectrometry tests and microscopy approaches.

The outcome

- A proven, published protocol for use within AstraZeneca's research and development activities.



Hogg K, Thomas J, Ashford D, et al. Quantification of proteins by flow cytometry: Quantification of human hepatic transporter P-gp and OATP1B1 using flow cytometry and mass spectrometry. *Methods*. 2015; 82: 38–46.



Case study

The user need

- Development and testing of a new imaging system.
- Provision of cross-comparison data.
- Access to various samples and application notes.

For this project, the Technology Facility acted as an expert partner, providing both specialist and forward-thinking conceptual consultancy advice. This was complemented with access to Category 2 wet labs, instrumentation and the ability to source a very wide range of sample types through the local academic user base. The Facility also provided an expert blend of staff, as well as both MSc and PhD student time, allowing exploitation and cross-comparison of multiple systems, all within the Facility's own purpose-built lab and desk space. Ultimately, this environment enabled a number of funding streams to be successfully leveraged. Once the new imaging system was developed, the Facility was also able to provide promotional support through application notes, academic publications, as well as external dissemination of information through invited talks and posters.

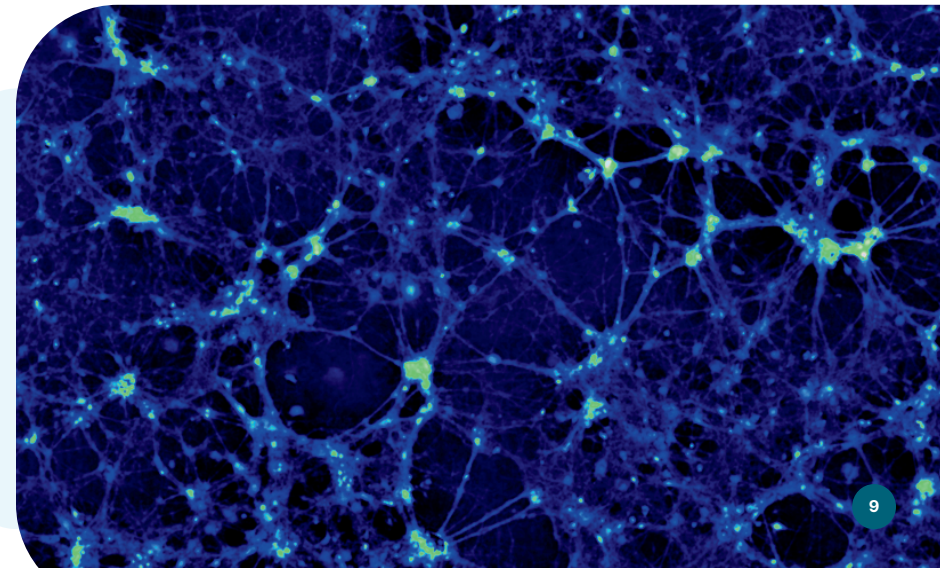
The outcome

- Creation of the successful commercial platform 'Livecyte'.
- Compilation of a host of biological application notes.
- Numerous academic publications.

The Facility is now home to two PhaseFocus application specialists. In turn, research teams at York have embraced the new technology developed with PhaseFocus, exploiting it for new fundamental research projects.

'The Facility's extraordinary team combine extremely deep technical and practical experience with a consistent willingness to go out of their way to help – no matter what the request. Their ever-present advice and practical assistance have been central to our continued commitment to renting space in York.'

Dr Martin Humphrey,
CEO Phasefocus Ltd, Sheffield



Genomics

The Genomics Laboratory provides expert support, advice, training and service provision for a wide range of genomic, epigenomic and transcriptomic applications.

Illumina sequencing

The Genomics Laboratory can provide end-to-end support, from experimental design through to analysis (working with members of the Data Science labs) for all your Illumina sequencing needs.

Nanopore long-read sequencing

The Genomics lab are experienced service providers of Oxford Nanopore Technologies sequencing, using both the MinION and PromethION systems. We have extensive experience working with a broad spectrum of sample types and species across a wide range of Nanopore sequencing applications.

Single-cell analysis

Close collaborations between the Genomics, Imaging & Cytometry and Data Science labs allow our facility to support state-of-the-art single cell isolation, processing and analysis.

Spatial sequencing

The Genomics, Imaging and Cytometry and Data Science labs work together to support a range of spatial analysis platforms as part of full service or collaborative projects, using both 10X Genomics and Nanostring systems.

Real-time (quantitative) PCR

The Genomics Laboratory offers access and training for qPCR, with three instruments supporting SYBR Green and TaqMan chemistries. All instruments can also be used for high-resolution melting studies.

Dr Sally James



Head of Genomics

 sally.james@york.ac.uk

 01904 328808

 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD

Sample quality analysis

We can perform rapid analysis of RNA and DNA quality and quantity using our Agilent 2100 Bioanalyser and 4200 Tapestation systems. Our Nanodrop Spectrophotometers support a wide range of analyses and allow for sample sizes as small as 1 µL.

Case study

The user need

- Long read DNA sequencing of 51 ash trees, to 30-fold genome coverage.

The Technology Facility's Genomics Laboratory was awarded a tender to work with Kew Gardens to generate high quality long-read genome sequence data for 51 individual ash trees in their collection.

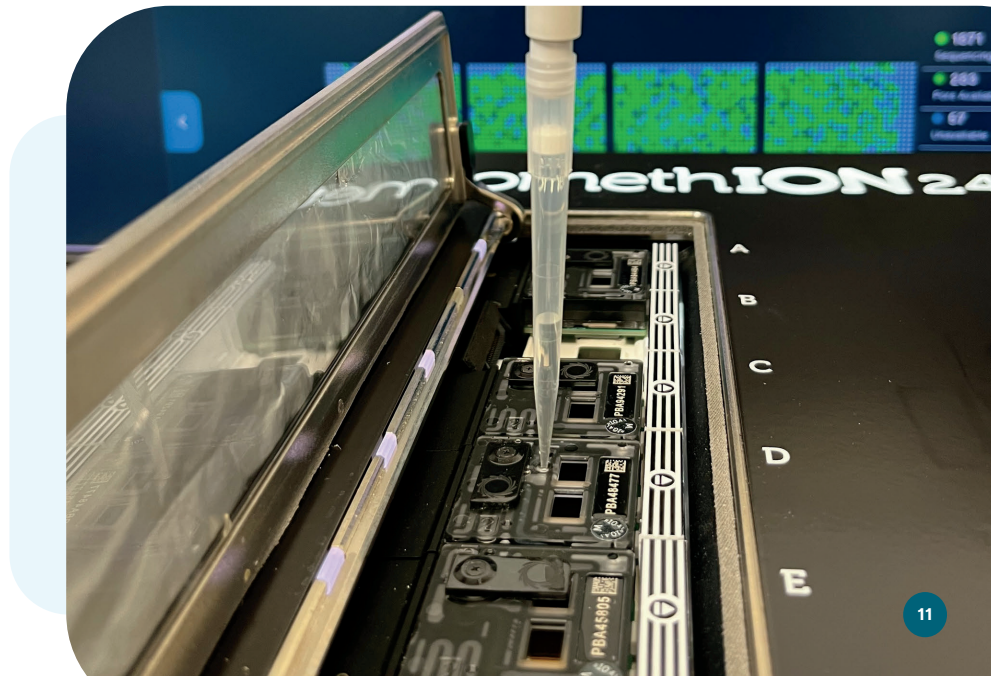
Facility staff acted as expert consultants on the DNA extraction process, providing advice and training to optimise our recommended extraction protocols for the team at Kew Gardens. High molecular weight DNA was successfully extracted from all samples, and a pipeline for this process was established for potential future work.

Long-read sequencing was then performed with Oxford Nanopore Technologies (ONT) PromethION sequencers and high-accuracy library preparation kits. Sequencing protocols were carefully tailored by Facility staff to meet (and often exceed) the specific requirements of the samples in question. The most recent versions of basecalling software were used to run super-accuracy basecalling (using our high-power computing capacity), with both raw and basecalled files transferred to the Kew Gardens team.

ONT long-read DNA sequencing platform is a rapidly evolving technology. Our Genomics Laboratory is recognised as a specialist in the field, able to utilise early-access sequencing reagents ahead of their general release, generating the highest quality read data possible.

The outcome

- Successful sequencing of all 51 samples submitted in the initial brief, with data delivered at or above the requested quality for all samples.
- Establishment of an efficient protocol for DNA extraction from plant leaf tissue with Kew Gardens, which they can now use in subsequent projects.



Metabolomics and Proteomics

We provide a range of mass spectrometry-based services using the comprehensive suite of instrumentation available in the Centre of Excellence in Mass Spectrometry.

Protein identification and sequencing

Our protein identification service has been running for over 20 years and is internationally known for providing reliable, high-confidence results with short turnaround times. We also offer a *de novo* sequencing service in cases where database matching with raw spectral data cannot identify a protein.

Proteomics

We offer a range of techniques for bottom-up proteomics, including label-free (both DIA and DDA) and label-based workflows. We work with whole cell samples across animal, plant, and microbial inputs, enriched samples (e.g. IP experiments), and specialised workflows including chemical crosslinking and proximity-based tagging approaches to help identify interacting partners.

Post-translational modifications (PTM)

We offer the following services for PTM identification and site mapping:

- Top-down sequencing by FT-ICR-MS
- T3 sequencing by ISD
- N- and C-terminal verification

Metabolomics and small-molecule services

We use combinations of GC-, LC-, and MALDI-MS techniques for non-targeted analysis, including profiling, annotation and relative quantification of small molecules including primary and secondary metabolites, drugs, xenobiotics, carbohydrates, and lipids.

We have a suite of software options for interrogation of mass spectrometry derived 'omic data, including Spectronaut, PEAKS StudioX-Pro, Byonic, Progenesis Q1, Compound and Proteome Discoverer, MetaboScape, and Mascot Distiller. We also design and implement bespoke workflows in KNIME and R. We are happy to discuss the best options to answer your biological questions and can advise on all aspects of experimental design.

Dr Tony Larson



Head of Metabolomics
and Proteomics

✉ tony.larson@york.ac.uk

☎ 01904 328733

📍 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD



Metabolomics and Proteomics in action

Case study

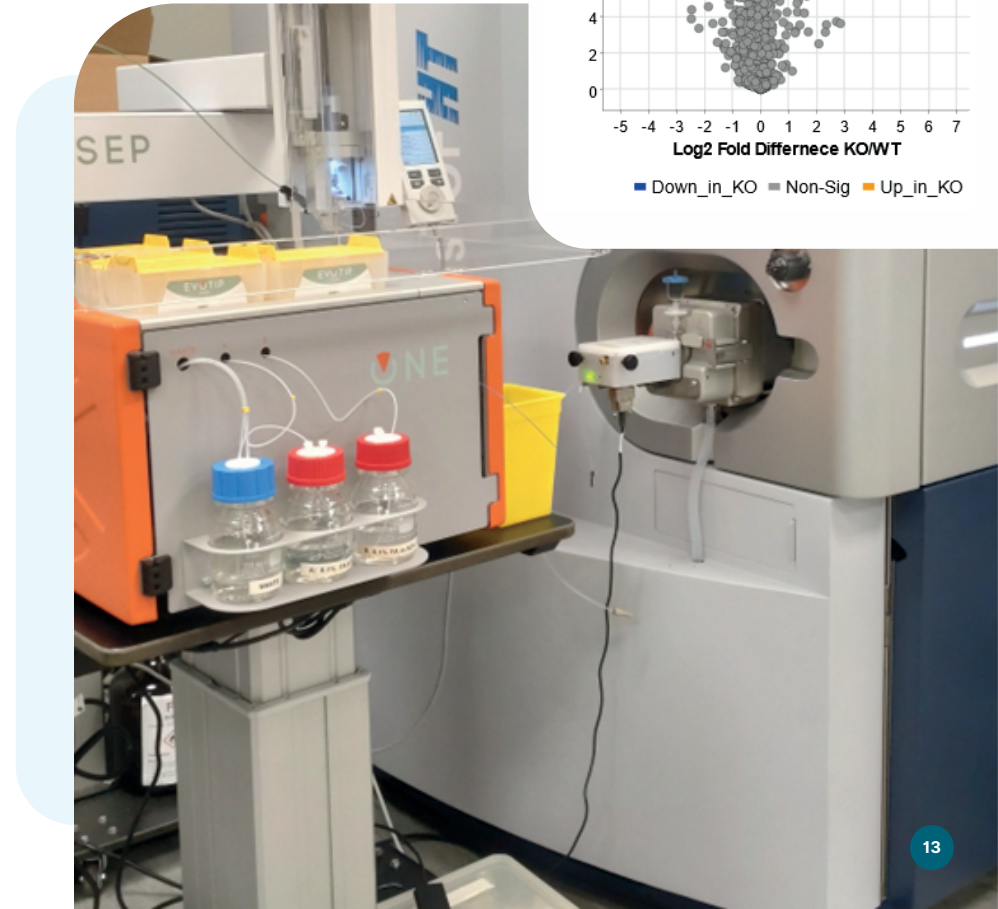
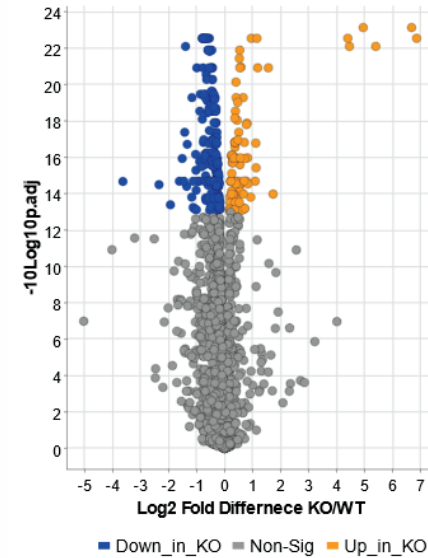
The user need

- Access to collaborative fee-for-service workflows for proteomic and metabolomic analyses.

In many Middle Eastern states capital investment in equipment is excellent but access to expertise is limited. Procurement of resources such as consumables in a timely manner to achieve project milestones can also be problematic. In this case, our experience in arranging workflows for external customers and our connection with the University of York legal contracts team enabled us to set the Technology Facility up as an official supplier on the Qatar Foundation supplier portal, giving researchers in universities across Qatar access to our services.

The outcome

We now regularly perform full-service contract proteomic and metabolomics workflows for a range of Qatari users, primarily supporting medical research workflows. Our solution includes consultation on experimental design, sample preparation, co-ordinating international sample shipping, sample analysis and data reporting.



Metabolomics and Proteomics in action

Case study

The user need

- Access to mass spectrometry services for protein identification and sequencing.

Many universities in the UK have access to pieces of equipment, such as mass spectrometers, that are utilised as part of specific research programmes but are not readily available for wider use. In some cases, the technical expertise may also not be on hand to support broader application.

The outcome

Since 2019, we have been engaging with the University of Nottingham, providing protein identification workflows with turnaround times of 1–2 weeks from sample receipt. This service has proven to be very desirable and successful and has become regularly utilised. The collaboration has led to the development of more complex LC-MS/MS-based proteomic workflows as well as several publications.



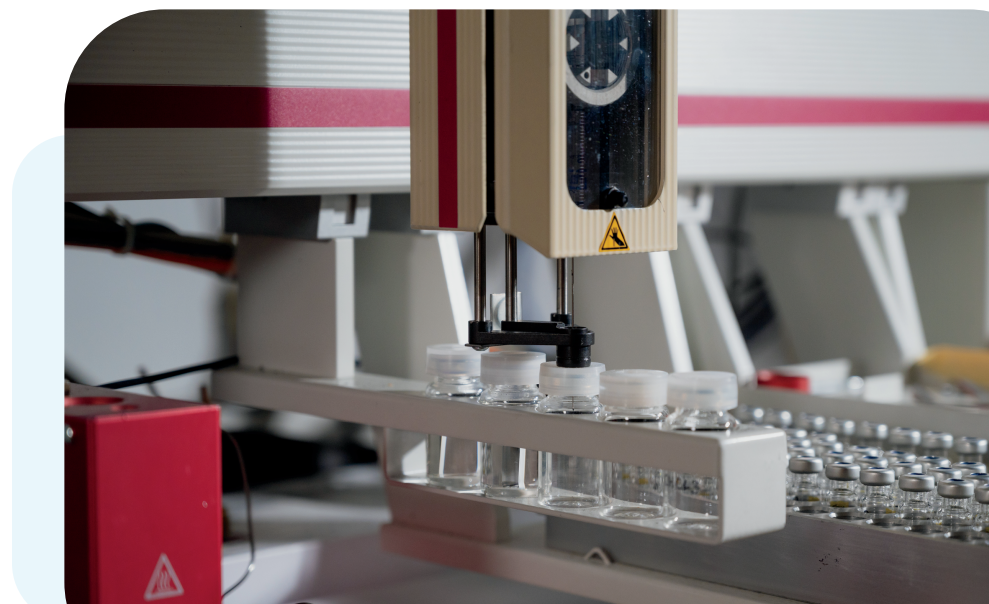
► Cells. 2023 Mar 16;12(6):912. doi: [10.3390/cells12060912](https://doi.org/10.3390/cells12060912)

Mitochondrial Haemoglobin Is Upregulated with Hypoxia in Skeletal Muscle and Has a Conserved Interaction with ATP Synthase and Inhibitory Factor 1

[Brad Ebanks](#)^{1,*}, [Gunjan Katyal](#)^{1,†}, [Chris Taylor](#)², [Adam Dowle](#)², [Chiara Papetti](#)³, [Magnus Lucassen](#)⁴, [Nicoleta Moisoi](#)⁵, [Lisa Chakrabarti](#)^{1,6,*}

Ebanks B, Katyal G, Taylor C, et al. Mitochondrial haemoglobin is upregulated with hypoxia in skeletal muscle and has a conserved interaction with ATP synthase and inhibitory factor 1.

Cells. 2023; 12: 912.



Molecular Interactions

Quantitative characterisation of macromolecules and their interactions is the remit of the Molecular Interactions Laboratory.

Protein properties, stability and interactions

The Molecular Interactions Laboratory offers a range of techniques for characterisation of biomacromolecules and their interactions, using biophysical and spectroscopic methods. We provide training and access to equipment but also advice on choice of method, experimental design and data analysis, and full service on submitted samples where appropriate. The lab has facilities for Surface Plasmon Resonance (Biacore SPR), Microscale Thermophoresis and Isothermal Titration Calorimetry to investigate and measure the interactions of proteins with potential ligands over a wide range of interaction strengths.

Macromolecule size, shape and stability

We have a Wyatt/Shimadzu SECMAALLS system and a Refeyn TwoMP mass photometer in regular use for solution molecular weight determination of proteins and other molecules, and a Beckman XL/I AUC system for sedimentation hydrodynamic studies. A Prometheus NanoDSF system provides simple, convenient protein QC but is also used for ligand binding as well as advanced stability assessments and an EI-Flex for single molecule FRET studies.

Spectroscopy

We have Circular Dichroism, UV-visible, fluorescence and single molecule FRET spectrometers with temperature control and other accessories for characterisation of macromolecules and small molecules, natural and synthetic, and a plate reader system capable of absorbance, fluorescence and luminescence assays in 96-well and other plate formats.

Dr Andrew Leech

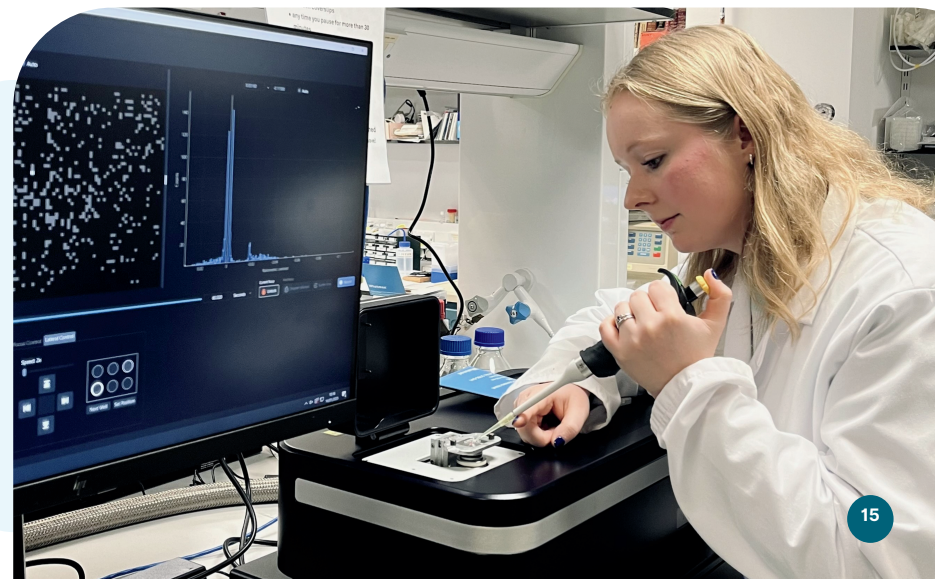


Head of Molecular
Interactions

 andrew.leech@york.ac.uk

 01904 328723

 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD



Protein Production

We offer a comprehensive service guided by a team of highly trained staff with extensive experience in the production of difficult-to-express recombinant proteins, particularly secreted enzymes and soluble domains of membrane-anchored targets.

Expression construct design and build

We are able to design, codon optimise and synthesise source genes, and sub-clone into an extensive TFPP and commercial vector library.

E. coli expression services

The lab is able to offer both cytoplasmic and periplasmic expression solutions, and provides high-throughput screening for soluble protein expression alongside an extensive range of scale-up capabilities.

K. phaffii expression services

We have access to novel, high-yielding expression vectors for secreted and folded protein expression, with a fully developed production pipeline (construct > fermentation > purification).

Insect and baculovirus expression services

We offer both intracellular and secreted expression pipelines, having developed scale-up and purification capabilities using bioreactors.

HEK293 expression services

We are also able to rapidly generate inducible, stable-transfected, suspension-adapted HEK293 cell lines, having developed scale-up and purification capabilities using bioreactors.

Dr Jared Cartwright



Head of Protein
Production

✉ jared.cartwright@york.ac.uk

☎ 01904 328797

📍 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD

Protein purification and characterisation services

Our lab has extensive experience in protein chemistry, affinity and conventional protein purification. Multi-dimensional chromatography options are available.

Fermentation and cell culture services

We utilise state-of-the-art aerobic and anaerobic facilities for fermentation processes, ranging from 0.5 to 50 litres in scale. We offer both batch and fed-batch cultivation methods. For cell culture applications, our capabilities extend from 0.5 to 10 litres. We support batch, fed-batch, and continuous cultivation techniques.

Protein Production in action

Case study

The user need

- Production of native extracellular receptor domains
- Full quality control (QC) of final recombinant proteins
- Supply to agreed specification

Protein expression from HEK293 (human embryonic kidney cells) is crucial when the post-translational modifications must be host-specific (for example, glycosylation patterns). Our system is expertly suited to the production of secreted and membrane-anchored targets.

Production at the Technology Facility is from inducible, stable-transfected cell lines that are prepared in suspension-adapted HEK293 cells. Advantages of this system include:

- Rapid preparation of stable cell lines (3 weeks)
- Tightly regulated expression providing consistent yields at any scale
- Serum-free media
- Defined scale-up process (bioreactors available)
- Defined affinity-purification procedures

Our expert staff, in collaboration with the Molecular Interactions lab, analysed the purified recombinant proteins and provided a full report including interpretation of the key findings. Examples of QC methods employed included peptide identification, intact mass spectrometry, thermal-shift assays, and determination of aggregation state by SEC-MALLS.

The outcome

We were able to produce a range of native extracellular receptor domains as high quality recombinant proteins meeting project specifications.

‘Thank you for the hard work that you and your team have put into generating our custom antigens. Your dedication and attention to detail is much appreciated. We have recommended your services to others looking into similar work.’

Sarah Greenhalgh,
Operations Director
GlycoScoreDX



Data Science

The Data Science Laboratory provides expertise and guidance across experimental design and data collection, through data analysis to downstream visualisation and dissemination.

Genomics and transcriptomics

We provide support to both external laboratories and other groups within the Technology Facility across a wide range of bioinformatics areas, including *de novo* genome assembly, SNP analysis, and RNA sequence analysis using short- and long-read sequencing technologies.

Single-cell and spatial analysis

We offer experimental design consultancy, processing and analysis for a range of single-cell and spatial analysis platforms.

Software design

We have extensive expertise in bespoke software design and implementation across multiple languages (R, Python and Rust), and can develop software solutions for multiple platforms including embedded systems, stand-alone computers and high-performance computing systems.

Computational pipeline services

The Data Science Laboratory maintains and runs a pipeline service that provides reliable, fast, and reproducible analyses for common workflows.

Training and tuition

We offer a wide range of training, from general programming and introductory courses to targeted training for specific subject fields. We also provide tailored tuition for individual laboratories.

Dr Alastair Droop

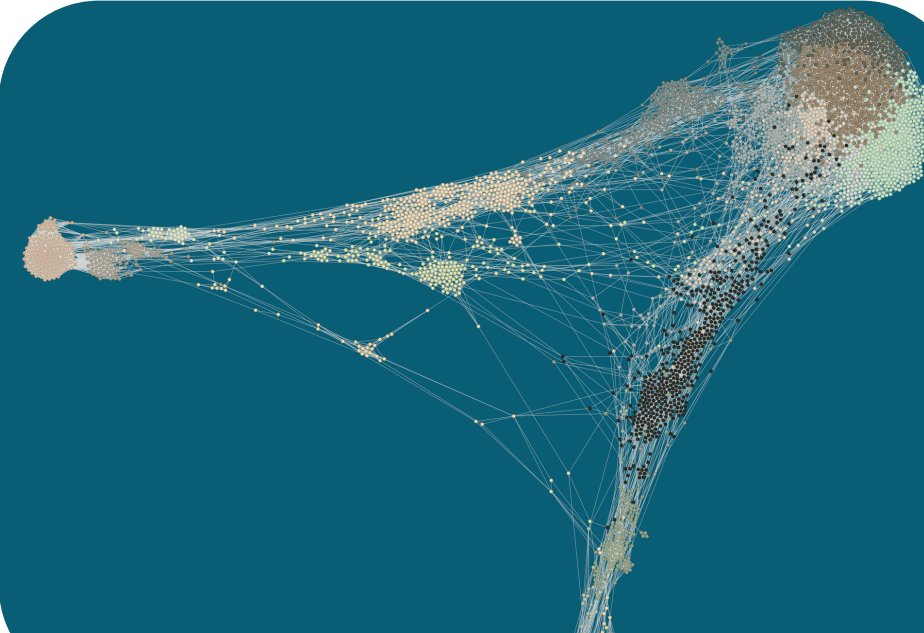


Head of Data Science

✉ alastair.droop@york.ac.uk

☎ 01904 328671

📍 Technology Facility,
Department of Biology,
University of York,
Wentworth Way, York,
YO10 5DD



Case study

The user need

- Development of software to support a new hyperspectral microscope system currently in development
- Both user-facing (front-end) and back-end software development

The user is developing a new imaging technique in collaboration with the Imaging and Cytometry lab in the Technology Facility. The Data Science group was approached to undertake the software development necessary to build the front-end software that would allow the users to control the new system. This development requires us to effectively implement the code that interfaces with the microscope hardware as well as providing a suitable “look and feel” for the users. Several new features were added to a preliminary LABview code, as this is an industry standard hardware interface development language.

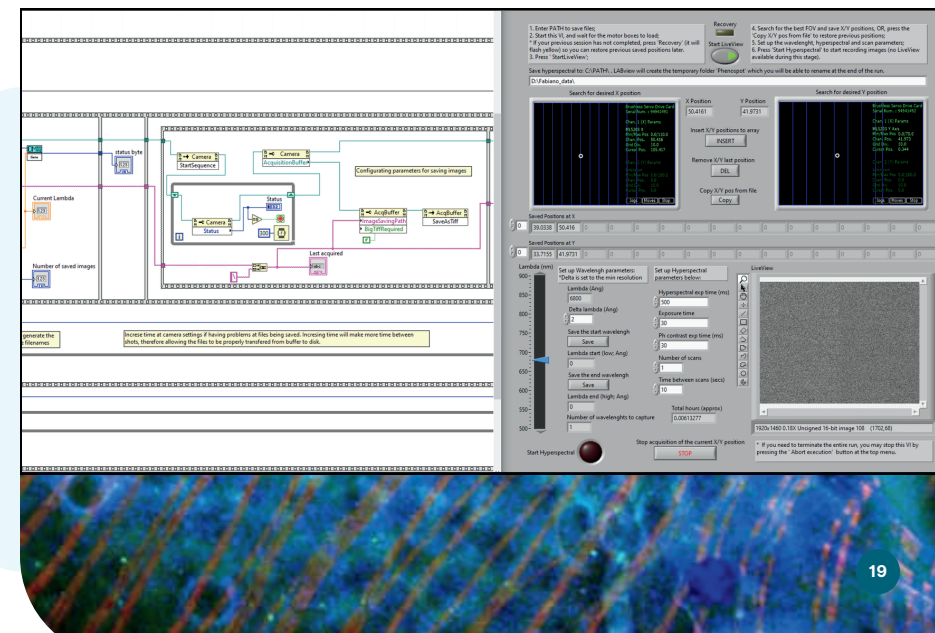
We were also tasked with developing high-performance back-end algorithms to improve the preprocessing speed of the system. This requires us to develop specific memory-safe and high-performance algorithms for data processing. We chose Rust for the back-end algorithm development due to its compromise between performance, memory safety and development speed.

The outcome

The microscope now has a performant front-end user interface that meets user requirements. We continue to work with users to add extra functionality as requested. Critically, we are able to complete data processing in a few seconds, down from hours in the original codebase.

‘The Data Science team were responsive and highly skilled. They engaged very well and really wanted to get to the bottom of the problem so they could provide the best possible service. The Technology Facility staff are willing and able to respond to all of our requested modifications.’

Professor Thomas Krauss



Our facility in numbers

Last year, our facility oversaw:



11

inter-lab
collaborations



24

involving
countries



60

resulting in
publications



30
staff

80+ high-end instruments
£500k+ equipment grants
per year

£45m+ active grants

supporting

400+
users

20 external academic
25 industrial groups

>200 internal and
100 external trainees,
with ~50% visiting
from overseas

every year, plus:

Partnerships with world-leading manufacturers:

- Zeiss
- Eluceda
- Mologic
- Carbonovia
- Beckman Coulter
- Oxford Nanopore Technologies
- Phase Focus
- Tomocube
- Royal Microscopical Society

Selected by UKRI
(BBSRC/MRC) as one of
only six exemplar nodes
for Euro-BioImaging



User testimony



Dr Sadaf Ashraf
University of Kent

The reason why I chose to work with the Technology Facility at the University of York is because I found the staff to be extremely friendly and helpful. I was particularly impressed with the expert advice they gave in the design of the workflow and the data analysis. Their commitment to supporting and training new external users through arranging online meetings, as well as offering to host them for visits to gain hands-on experience, illustrates their passion for providing and nurturing a collaborative research environment.



Dr Debbie Stokes
Founder of NANOVIIZ

The infrastructure of the Facility makes it very easy for external users to apply for and gain access to a broad range of specialised equipment, in order to produce high-quality, complementary results. Further, the staff's support in designing experiments, developing methodologies and discussing/analysing results was immensely valuable.



Ryan Mathew FRCS (NeuroSurg.) PhD
University of Leeds

The team both understood our needs, as well as how to adapt their instrumentation to find the optimised approach.

The Nanostring DSP is also an extremely specialised and expensive piece of equipment which requires dedicated expert technical input. There are very few examples of this equipment which can be accessed elsewhere and York are enabling this with ease.



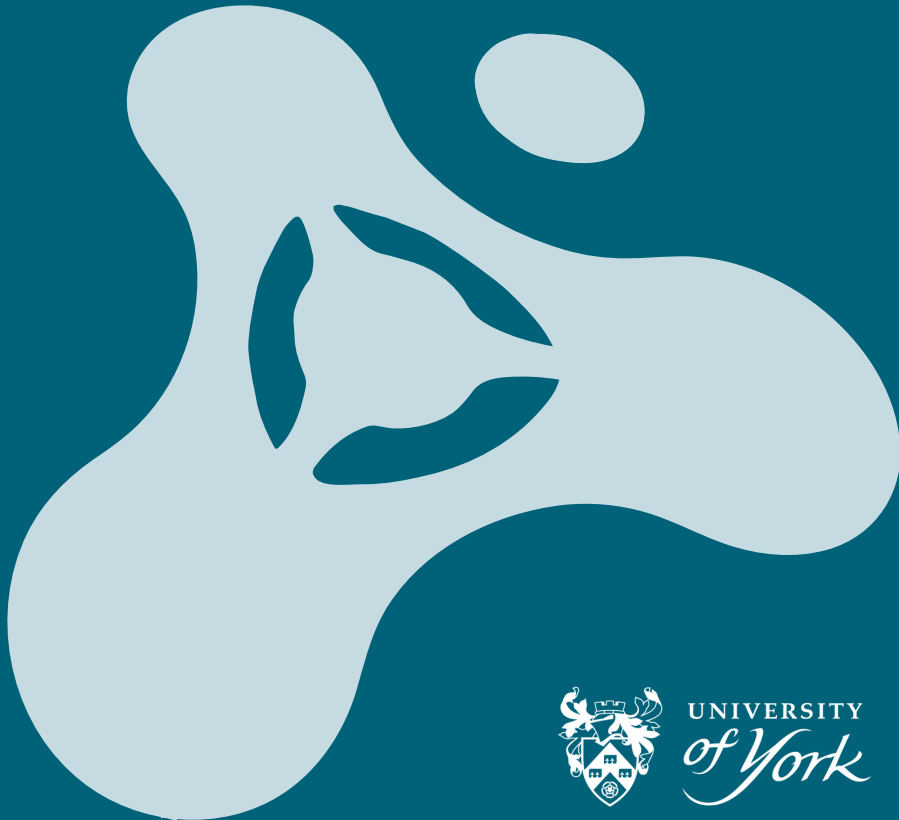
Dr. Omar Muhammad Khan
Hamad Bin Khalifa University

My lab had the pleasure of working with the BTF-MAP lab for several proteomics projects, I cannot recommend their services highly enough. They exceeded my expectations taking the time to understand the requirements of my projects, provided expert feedback, and incorporated advanced bioinformatics tools for the data analysis. One aspect that truly sets the BTF-MAP lab apart is its customer service. Communication was clear, transparent, and timely. The staff were friendly, professional, and truly invested in the success of my project. The turnaround time was also much quicker than my expectations.

Notes



Bioscience Technology Facility



Get in contact

+44 (0)1904 328821

btf-enquiries@york.ac.uk

Find out more at:

www.york.ac.uk/biology/technology-facility/

Like and follow us:



@YorkBioscience



York Bioscience



@yorkbioscience.bsky.social

